



# Comparison of aromatic monomers in lignocellulosic biomass prehydrolysates

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Differences in the relative toxicity of xylose-rich prehydrolysates derived from woody and herbaceous feedstocks are likely due to the relative abundance of a variety of inhibitory compounds. Acetate, as well as several aromatic monomers, has been shown to be an inhibitor of the xylose-fermenting yeast, *Pichia stipitis*. Comparative information on the concentration of known and likely inhibitors, other than acetate, is lacking. The present study provides data on the aromatic monomer composition of representative herbaceous and woody prehydrolysates. Dilute-acid prehydrolysates were prepared from three feedstocks; two herbaceous, corn stover and switchgrass (*Panicum virgatum* L.), and one woody (poplar). The prehydrolysates were neutralized with Ca(OH)<sub>2</sub>, extracted with ethyl acetate, trimethylsilylated, and analyzed by GC-MS. Fourteen aromatic monomers were tentatively identified by comparison with published mass spectra. The concentrations of the aromatic monomers totalled 112, 141 and 247 mg L<sup>-1</sup> for corn stover, switchgrass and poplar prehydrolysates, respectively. This is also the order of increasing inhibition of growth and ethanol productivity observed for *Pichia* fermentations. The woody prehydrolysate contained approximately four-fold more syringyl-based monomers than did the herbaceous prehydrolysates, while guaiacyl-containing compounds were more evenly distributed.

**Keywords:** lignocellulosic biomass; prehydrolysate; *Pichia stipitis*; aromatic; inhibitors

## Introduction

The production of fuel ethanol from lignocellulosic biomass feedstocks has several benefits including domestic availability, pollution reduction and ease of introduction into existing gasoline and diesel distribution networks [12,25]. One widely studied process for converting lignocellulosic biomass to ethanol involves a pretreatment of the feedstock with dilute acid (approximately 1.0% (w/w) sulfuric acid) at temperatures over 140°C, conditions which catalyze the hydrolysis of biomass hemicellulose, but leave the cellulose fraction largely intact [17]. This dilute-acid pretreatment yields a solids stream, consisting of mostly lignin and cellulose, and an aqueous stream, called the prehydrolysate, which contains hemicellulose-derived xylose, lesser amounts of other carbohydrates, sugar degradation products, lignin degradation products, acetic acid, and other compounds [15]. Sugars (mostly xylose) in the prehydrolysate typically represent 15–30% of the original dry weight of the biomass [7]. Rapid and efficient fermentation of these sugars is essential for making biomass-to-ethanol conversion processes economically viable [25].

Dilute-acid prehydrolysates contain many compounds other than sugars, and some of these inhibit fermentation by and growth of microorganisms. These inhibitors can be divided into groups based on their origin [15]: (1) compounds released during pretreatment, eg acetic acid;

(2) sugar degradation products, eg furfural; (3) lignin degradation products, eg syringaldehyde; (4) fermentation products, eg ethanol; and (5) contaminants released by processing equipment, eg chromium. For a review of prehydrolysate fermentation and a discussion of the role of inhibitors, see Olsson and Hahn-Hägerdahl [15].

Several papers describing ethanol yields from *Pichia stipitis* fermentations of dilute-acid prepared lignocellulosic prehydrolysates have been published, including studies of prehydrolysate derived from aspen wood [26], red oak [22], sugar cane bagasse [20], corn cob [10], mixed wood chips [18], eucalyptus [9], switchgrass [8], corn stover [8,16], poplar [8,16], rice straw [13], *Pinus radiata* [19] and wheat straw [4]. *Pichia*, although an efficient fermenter of xylose is susceptible to inhibition by toxic compounds in lignocellulosic prehydrolysates, leading to lower ethanol productivities (g ethanol L<sup>-1</sup> h<sup>-1</sup>) and yields (g ethanol g sugar consumed<sup>-1</sup>) compared to those achieved in fermentations of control media [15]. Among the compounds identified in prehydrolysates which are known to inhibit *Pichia* fermentations are vanillin, syringaldehyde, acetic acid and furfural [5,22,23]. The lignin-derived compounds, ie syringaldehyde and vanillin, have been shown to be particularly potent inhibitors of ethanol production and cell growth [5,22], especially when compared on a concentration basis to acetic acid or sugar degradation compounds. In particular, vanillin, a guaiacyl-containing compound, was shown to be more inhibitory than representative syringyl- and hydroxybenzyl-containing compounds [5].

Phenolic compounds in prehydrolysates prepared from woody substrates have been described [1,3,22,26]. In contrast, relatively little is known about the lignin-derived compounds in prehydrolysate obtained from herbaceous

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This is Oregon State University Agricultural Experiment Station Technical Publication Number 11330

Received 24 April 1998; accepted 8 June 1998

lignocellulosics such as switchgrass (*Panicum virgatum* L.) and corn stover. Switchgrass and corn stover are considered to be excellent candidate feedstocks for conversion to ethanol [6,21], and thus are of particular interest.

The aim of the present study was to identify and quantify phenolics and lignin-derived compounds found in the prehydrolysate derived from two herbaceous feedstocks (switchgrass and corn stover) and one woody feedstock (poplar). These feedstocks were chosen based on their potential for use in biomass-to-ethanol processes and because the relative fermentability of their prehydrolysates has been established [8]. The results provide analytical information that should be useful in determining the chemical basis for differences in rates and extents of prehydrolysate fermentations as well as for designing detoxification processes [15].

## Materials and methods

### Sample preparation

Milled and dried feedstocks (poplar, corn stover and switchgrass) were treated in a 0.6-L stainless steel Parr reactor at 10% solids (180°C, 1% (w/w) H<sub>2</sub>SO<sub>4</sub>, 1 min) as described [7]. The resulting material was filtered through VWR 413 grade paper, and neutralized to pH 6.0 with Ca(OH)<sub>2</sub> as described [8]. The pretreatment conditions were chosen to optimize the yield of xylose in the liquid prehydrolysate. One milliliter of 0.13 g L<sup>-1</sup> *o*-vanillin (Sigma, St Louis, MO, USA) internal standard solution was added to 10 ml of prehydrolysate. Ten milliliters of ethyl acetate were added and mixed by inversion. After separation of the two phases, the upper layer was transferred to a test tube and dried under N<sub>2</sub> at 40°C for 15 min. The residue was redissolved in 0.5 ml of ethyl acetate and transferred to a vial to which was added 50 µl pyridine and 300 µl of bis(trimethylsilyl)trifluoroacetamide (BSTFA) from Sigma. The mixture was allowed to react at room temperature overnight.

### GC-MS

Gas chromatography/mass spectrometry was performed on a 10-m SE-54 capillary column using a Finnigan GC/MS. Mass spectra were obtained at 70 eV. For quantification of tentatively identified phenolic compounds, a 1 : 1 response ratio with *o*-vanillin was assumed.

## Results/discussion

Partial total ion count (TIC) chromatograms obtained from TMS-derivatised ethyl acetate extracts from corn stover, switchgrass and poplar prehydrolysates are shown in Figure 1. In all cases, chromatograms reflect neutralized prehydrolysates, ie suitable for fermentation. This approach was taken to facilitate comparison of the GC-MS data with fermentation results. Ethyl acetate was used for extraction because it has been shown to remove phenolic compounds such as parahydroxybenzoic acid and vanillin from aspen prehydrolysates. Ethyl acetate was also shown to reduce inhibition of microbial growth and fermentation of prehydrolysates [26]. Thus it can be expected that microbial

inhibitors, along with non-inhibitory compounds, are to be found in an ethyl acetate extract.

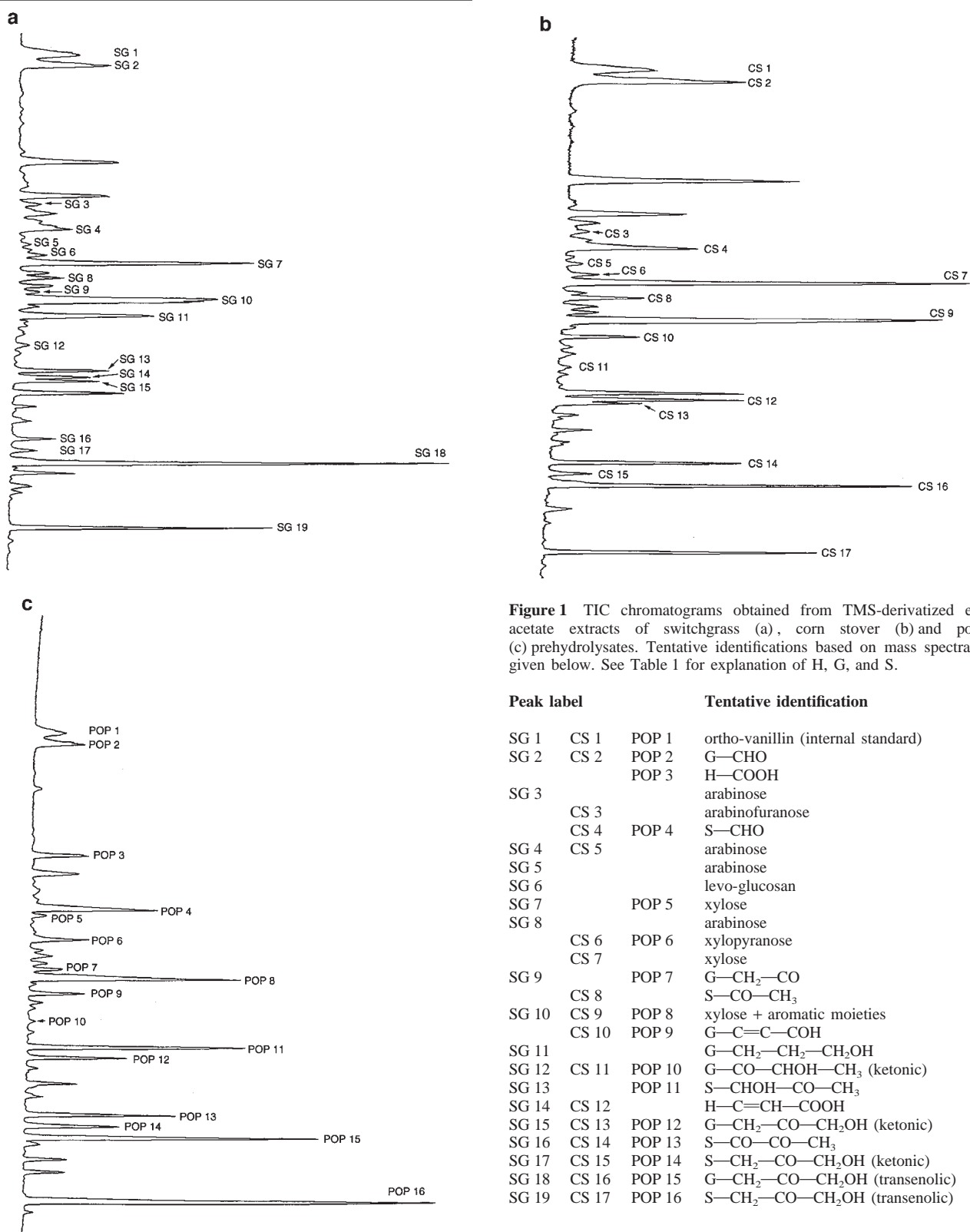
By matching mass spectra with those in the literature [11,14,27], 14 compounds with structures indicative of lignin monomers or esterified phenolics were tentatively identified. Table 1 summarizes the identification and quantification of the chromatogram peaks from the three feedstocks. The concentrations of the identified lignin-derived components total 247, 141 and 112 mg L<sup>-1</sup> for poplar, switchgrass and corn stover prehydrolysates, respectively. This is also the order of decreasing microbial inhibition, as measured by ethanol yield, productivity, and cell mass increase in *Pichia* fermentations, as shown in Table 2. In addition, this is also the order of decreasing Klason lignin content for the feedstocks [7]. Similarly, the Klason lignin content of native woody feedstocks was found to be negatively associated with ethanol yields using *Saccharomyces* in simultaneous saccharification and fermentation experiments [24]. The acetate concentrations averaged 1.3, 2.0 and 2.9 g L<sup>-1</sup> for corn stover, switchgrass and poplar prehydrolysates, respectively. Furfural and hydroxymethylfurfural concentrations were less than 0.01 g L<sup>-1</sup> for all neutralized prehydrolysates.

The most striking difference in both the type and amount of aromatic compounds was found among the syringyl compounds. In poplar prehydrolysate, the concentration of syringyl-containing compounds was approximately four-fold higher than in switchgrass or corn stover prehydrolysates. The guaiacyl compounds were more evenly distributed among the three prehydrolysates, with corn stover having the lowest concentration of identified guaiacyl derivatives, and switchgrass the highest. Although other work indicates that guaiacyl-containing monomers are highly toxic to *Pichia* [5], the concentrations used in that study were far higher than anything seen in the prehydrolysates examined here. The work currently described was conducted on native, unextracted biomass, so it is difficult to determine whether differences in the ratios of the three main phenyl derivatives are due to differences in the core lignin structures of the three feedstocks, or if they represent differences in more-easily solubilized non-core lignin materials.

The concentrations of syringaldehyde (S-CHO, Table 1) and vanillin (G-CHO, Table 1) in the poplar prehydrolysate are similar to those reported by Buchert *et al* [3] for birch wood prehydrolysate prepared by steam treatment and extracted with dimethylchloride without neutralization, although the concentrations of 1-guaiacyl acetol (G-CH<sub>2</sub>-CO-CH<sub>2</sub>OH, Table 1) and 1-syringyl acetol (S-CH<sub>2</sub>-CO-CH<sub>2</sub>OH, Table 1) in poplar prehydrolysate were found to be 20–30 fold higher. In comparison to red oak prehydrolysate pretreated under similar conditions [22], poplar prehydrolysate contained 5-fold less vanillin and 10-fold less syringaldehyde, although the extraction and preparation regimens were different.

Compounds which were unique to specific prehydrolysates included vanillyl propanol (G-CH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>OH, Table 1) in switchgrass, syringyl methyl ketone (S-CO-CH<sub>3</sub>, Table 1) in corn stover and para-hydroxybenzoic acid (H-COOH, Table 1) in poplar. Para-hydroxybenzoic acid is a major component in poplar-derived hydrolysates [1,2].

In previous work [8] it was shown that prehydrolysates



**Figure 1** TIC chromatograms obtained from TMS-derivatized ethyl acetate extracts of switchgrass (a), corn stover (b) and poplar (c) prehydrolysates. Tentative identifications based on mass spectra are given below. See Table 1 for explanation of H, G, and S.

Peak label	Tentative identification
SG 1	POP 1 ortho-vanillin (internal standard)
SG 2	POP 2 G—CHO
	POP 3 H—COOH
SG 3	arabinose
	CS 3 arabinofuranose
SG 4	CS 4 POP 4 S—CHO
SG 5	CS 5 arabinose
SG 6	arabinose
SG 7	POP 5 levo-glucosan
SG 8	POP 5 xylose
	arabinose
	CS 6 POP 6 xylopyranose
	CS 7 xylose
SG 9	POP 7 G—CH <sub>2</sub> —CO
	CS 8 S—CO—CH <sub>3</sub>
SG 10	CS 9 POP 8 xylose + aromatic moieties
	CS 10 POP 9 G—C=C—COH
SG 11	G—CH <sub>2</sub> —CH <sub>2</sub> —CH <sub>2</sub> OH
SG 12	CS 11 POP 10 G—CO—CHOH—CH <sub>3</sub> (ketonic)
SG 13	POP 11 S—CHOH—CO—CH <sub>3</sub>
SG 14	CS 12 H—C=CH—COOH
SG 15	CS 13 POP 12 G—CH <sub>2</sub> —CO—CH <sub>2</sub> OH (ketonic)
SG 16	CS 14 POP 13 S—CO—CO—CH <sub>3</sub>
SG 17	CS 15 POP 14 S—CH <sub>2</sub> —CO—CH <sub>2</sub> OH (ketonic)
SG 18	CS 16 POP 15 G—CH <sub>2</sub> —CO—CH <sub>2</sub> OH (transenolic)
SG 19	CS 17 POP 16 S—CH <sub>2</sub> —CO—CH <sub>2</sub> OH (transenolic)

from herbaceous feedstocks were more easily fermentable than those from woody feedstocks, and that the discrepancies could be only partially attributed to acetate concentration differences. It is demonstrated here that prehydrolys-

ates derived from herbaceous feedstocks (switchgrass and corn stover) contain much less phenolic monomers than poplar-derived prehydrolysate. When considered alongside the lower acetate concentrations of the herbaceous prehy-

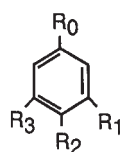
**Table 1** Lignin-derived compounds tentatively identified in dilute-acid prehydrolysates prepared from poplar, switchgrass and corn stover

Phenolic moiety <sup>c</sup>	R <sub>0</sub>	Chromatogram peak labels	Poplar (mg L <sup>-1</sup> )	Switchgrass (mg L <sup>-1</sup> )	Corn stover (mg L <sup>-1</sup> )
H	—COOH	POP3	11.1 (0.8) <sup>a</sup>	nd <sup>b</sup>	nd
H	—C=CH—COOH	SG14, CS13	nd	7.7 (0.2)	10.9 (0.2)
S	—CHO	CS4, POP4	29.3 (2.0)	nd	10.0 (0.3)
S	—CHOH—CO—CH <sub>3</sub>	SG13, POP11	35.3 (2.2)	9.1 (0.3)	10.0 (0.6)
S	—CH <sub>2</sub> —CO—CH <sub>2</sub> OH	SG17,19; CS15,17; POP14, 16	74.4 (4.0)	26.3 (1.5)	17.2 (0.8)
S	—CO—CH <sub>3</sub>	CS8	nd	nd	5.2 (0.1)
S	—CO—CO—CH <sub>3</sub>	SG16, CS14, POP13	17.0 (0.8)	2.6 (0.1)	9.5 (0.3)
G	—CO—CHOH—CH <sub>3</sub>	SG12, CS11, POP10	1.0 (0.1)	2.0 (0.1)	nd
G	—CHO	CS2, SG2, POP2	15.1 (0.4)	16.5 (0.6)	19.6 (0.2)
G	—CH <sub>2</sub> —CO	SG9, POP7	4.4 (0.4)	1.5 (0.1)	nd
G	—CH <sub>2</sub> —CH <sub>2</sub> —CH <sub>2</sub> OH	SG11	nd	18.4 (0.8)	nd
G	—C=CH—COH	CS10, POP9	8.5 (0.5)	trace	4.0 (0.6)
G	—CH <sub>2</sub> —CO—CH <sub>2</sub> OH	SG15, 18; CS13, 16; POP12, 15	51.2 (2.5)	56.7 (3.0)	26.0 (1.1)
Total			247.3	140.8	112.4

<sup>a</sup>Values in parentheses are standard errors of the means.

<sup>b</sup>nd = not detected.

<sup>c</sup>



Moiety	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>
Hydroxyphenyl (H)	H	OH	H
Guaiacyl (G)	OCH <sub>3</sub>	OH	H
Syringyl (S)	OCH <sub>3</sub>	OH	OCH <sub>3</sub>

**Table 2** Inhibitor concentrations and fermentation results obtained from poplar, switchgrass and corn stover prehydrolysates

Prehydrolysate source	<i>p</i> -Hydroxy-phenyl monomers (mg L <sup>-1</sup> )	Guaiacyl monomers (mg L <sup>-1</sup> )	Syringyl monomers (mg L <sup>-1</sup> )	Acetate <sup>a</sup> (g L <sup>-1</sup> )	Ethanol yield <sup>a</sup> (g ethanol g sugar <sup>-1</sup> )	Ethanol productivity <sup>a</sup> (g ethanol L <sup>-1</sup> h <sup>-1</sup> )	Final dry cell mass <sup>a</sup> (relative to control)
Poplar	11.1	80.2	156.0	2.9	0.34	0.25	-15%
Switchgrass	7.7	95.1	38.0	2.0	0.38	0.54	+5%
Corn stover	10.9	49.6	51.9	1.3	0.41	0.79	+7%

<sup>a</sup>Calculated from Reference [8].

drolysates, the evidence presented here provides a good rationale for the lower toxicity of herbaceous prehydrolysates. Whether this trend can be generalized to other feedstocks can only be determined by direct comparison. These data provide a basis for comparison in assessing the feasibility of feedstocks for biomass-to-ethanol processes.

## Acknowledgements

This work was funded in part by the National Renewable Energy Laboratory, Golden, Colorado.

## References

- Ando S, I Arai, K Kiyoto and S Hanai. 1986. Identification of aromatic monomers in steam-exploded poplar and their influences on ethanol fermentation by *Saccharomyces cerevisiae*. *J Ferment Technol* 64: 567–570.
- Bardet M and DR Robert. 1985. On the reactions and degradation of the lignin during steam hydrolysis of aspen wood. *Svensk Papperst 6*: 61–67.
- Buchert J, K Niemelä, J Puls and K Poutanen. 1990. Improvement in the fermentability of steamed hemicellulose hydrolysate by ion exclusion. *Process Biochem Int* 25: 176–180.
- Delgenes J, R Moletta and J Navarro. 1988. Acid hydrolysis of wheat straw and process considerations for ethanol fermentation by *Pichia stipitis* Y7124. *Process Biochem Int* 12: 132–135.
- Delgenes JP, R Moletta and JM Navarro. 1996. Effects of lignocellulosic degradation products on ethanol fermentations of glucose and xylose by *Saccharomyces cerevisiae*, *Zymomonas mobilis*, *Pichia stipitis* and *Candida shehatae*. *Enzyme Microb Technol* 19: 220–225.
- Downing M, S McLaughlin and MJ Walsh. 1995. In: Second Biomass Conference of the Americas: Energy, Environment, Agriculture, and Industry Proceedings. National Renewable Energy Laboratory, Golden, CO.
- Esteghlalian A, AG Hashimoto, JJ Fenske and MH Penner. 1997. Modeling and optimization of the dilute-acid pretreatment of corn stover, poplar and switchgrass. *Biores Technol* 59: 129–136.
- Fenske JJ, AH Hashimoto and MH Penner. Relative fermentability of lignocellulosic dilute-acid prehydrolysates: application of a *Pichia stipitis*-based toxicity assay. *Appl Biochem Biotechnol* (in press).
- Ferrari MD, E Neirotti, C Albornoz and E Saucedo. 1992. Ethanol production from eucalyptus wood hemicellulose hydrolysate by *Pichia stipitis*. *Biotechnol Bioeng* 40: 753–759.
- Hahn-Hägerdahl B, H Jeppsson, L Olsson and A Mohagheghi. 1994. An interlaboratory comparison of the performance of ethanol-producing micro-organisms in a xylose-rich acid hydrolysate. *Appl Microbiol Biotechnol* 41: 62–72.
- Lapierre C, C Rolando and B Monties. 1983. Characterization of poplar lignins acidolysis products: capillary gas-liquid and liquid-liquid chromatography of monomeric compounds. *Holzforsch* 37: 189–198.

- 12 Lynd LR, JH Cushman, RJ Nichols and CE Wyman. 1991. Fuel Ethanol from cellulosic biomass. *Science* 251: 1318–1323.
- 13 Moniruzzaman M. 1995. Alcohol fermentation of enzymatic hydrolysate of exploded rice straw by *Pichia stipitis*. *World J Microbiol Biotech* 11: 646–648.
- 14 Niemelä K and E Sjöström. 1986. Simultaneous identification of aromatic and aliphatic low molecular weight compounds from alkaline pulping liquor by capillary gas-liquid chromatography-mass spectroscopy. *Holzforsch* 40: 361–368.
- 15 Olsson L and B Hahn-Hägerdahl. 1996. Fermentation of lignocellulosic hydrolysates for ethanol production. *Enzyme Microb Technol* 18: 312–331.
- 16 Parekh SR, RS Parekh and M Wayman. 1988. Ethanol and butanol production by fermentation of enzymatically saccharified SO<sub>2</sub>-prehydrolyzed lignocellulosics. *Enz Microb Tech* 10: 660–668.
- 17 Penner MH, AG Hashimoto, A Esteghlalian and JJ Fenske. 1996. In: *Agricultural Materials as Renewable Resources—Nonfood and Industrial Applications* (Fuller G, TA McKeon and DD Bills, eds), pp 12–31, American Chemical Society, Washington DC, USA.
- 18 Perego P, A Converti, E Palazzi, M del Borghi and G Ferraiolo. 1990. Fermentation of hardwood hemicellulose hydrolysate by *Pachysolen tannophilus*, *Candida shehatae* and *Pichia stipitis*. *J Ind Microbiol* 6: 157–164.
- 19 Qureshi N and GJ Manderson. 1991. Ethanol production from sulphuric acid wood hydrolysate of *Pinus radiata* using free and immobilized cells of *Pichia stipitis*. *J Ind Microbiol* 7: 117–122.
- 20 Roberto IC, LS Lacis, MFS Barbosa and IM de Mancilha. 1991. Utilization of sugar cane bagasse hemicellulosic hydrolysate by *Pichia stipitis* for the production of ethanol. *Process Biochem* 26: 15–21.
- 21 Sanderson MA, RL Reed, SB McLaughlin, SD Wullschleger, BV Conger, DJ Parrish, DD Wolf, C Taliaferro, AA Hopkins, WR Ocumpaugh, MA Hussey, JC Read and CR Tischler. 1996. Switchgrass as a sustainable bioenergy crop. *Bioresource Technol* 56: 83–93.
- 22 Tran AV and RP Chambers. 1986. Ethanol fermentation of red oak acid prehydrolysate by the yeast *Pichia stipitis* CBS 5776. *Enzyme Microb Technol* 8: 439–444.
- 23 van Zyl C, BA Prior and JC du Preez. 1991. Acetic acid inhibition of d-xylose fermentation by *Pichia stipitis*. *Enzyme Microb Technol* 13: 82–86.
- 24 Vinzant TB, CI Ehrman, WS Adney, SR Thomas and ME Himmel. 1997. Simultaneous saccharification and fermentation of pretreated hardwoods. Effect of native lignin content. *Appl Biochem Biotechnol* 62: 99–104.
- 25 von Sivers M and G Zacchi. 1996. Ethanol from lignocellulosics: a review of the economy. *Bioresource Technol* 56: 131–140.
- 26 Wilson JJ, L Deschatelets and NK Nishikawa. 1989. Comparative fermentability of enzymatic and acid hydrolysates of steam-pretreated aspenwood hemicellulose by *Pichia stipitis* CBS 5776. *Appl Microbiol Biotechnol* 31: 592–596.
- 27 Wittkowski R. 1985. *Phenole im Räucherrauch-Nachweis und Identifizierung*. VCH Verlagsgesellschaft mbH, Weinheim, Germany.